

Example of microplastic analysis

Natalja Buhhalke

Tallinn University of Technology

Department of Marine Systems



**Before you
begin**

Know your target



It determines ALL your
next steps



Where from this sample was taken

LIQUID *seawater, freshwater, wastewater*

SOLID *sediment, soil, sludge*

BIOTA *algae, insects, fish, etcetera*

- *surface? subsurface? Above ocean floor? Before settling tank? before primary treatment?*
- *homogenised? **LOCATION** depth? top layer? low thermodynamic flow? Overturn depth?*
- *Pristine population? juvenile? contaminated area? lab-grown? DGI tract only?*

Sample treatment

- To use as less steps as possible

MethodsX 9 (2022) 101603



Contents lists available at ScienceDirect

MethodsX

journal homepage: www.elsevier.com/locate/mex



Method Article

Hide-and-seek: Threshold values and contribution towards better understanding of recovery rate in microplastic research



Inta Dimante-Deimantovica^{a,b,*}, Natalija Suhareva^a, Marta Barone^{a,c}, Ieva Putna-Nimane^a, Juris Aigars^a

^aLatvian Institute of Aquatic Ecology, Agency of Daugavpils University, 4 Voleru Str., Riga LV-1007, Latvia

^bInstitute of Biology, University of Latvia, 1 Jelgavas Str., Riga LV-1004, Latvia

^cThe Faculty of Natural Sciences and Mathematics, Daugavpils University, 1 Parades Str., Daugavpils LV-5401, Latvia

ABSTRACT

Microplastic pollution has become one of the most pressing environmental issues. A fundamental criterion for risk assessment is the concentration of found microplastic that can be altered during microplastic isolating from the sample. Recovery rate (i.e. positive control) is an important feedback component that identifies accuracy, quality and efficiency of sample processing, same as physical and chemical impact. Here, using 100 µm red polystyrene (PS) beads we have tested some methodological steps that can be responsible for the possible microplastic losses during sample treatment and based on that, we provided a recovery rate threshold values. Our results support that the choice of the extraction method (vacuum filtration versus wet sieving) results in lower recoverability when vacuum filtration is used and that used separatory funnels size versus material amount impacts the efficiency or recoverability in density separation. We have also analysed microplastic recovery rate

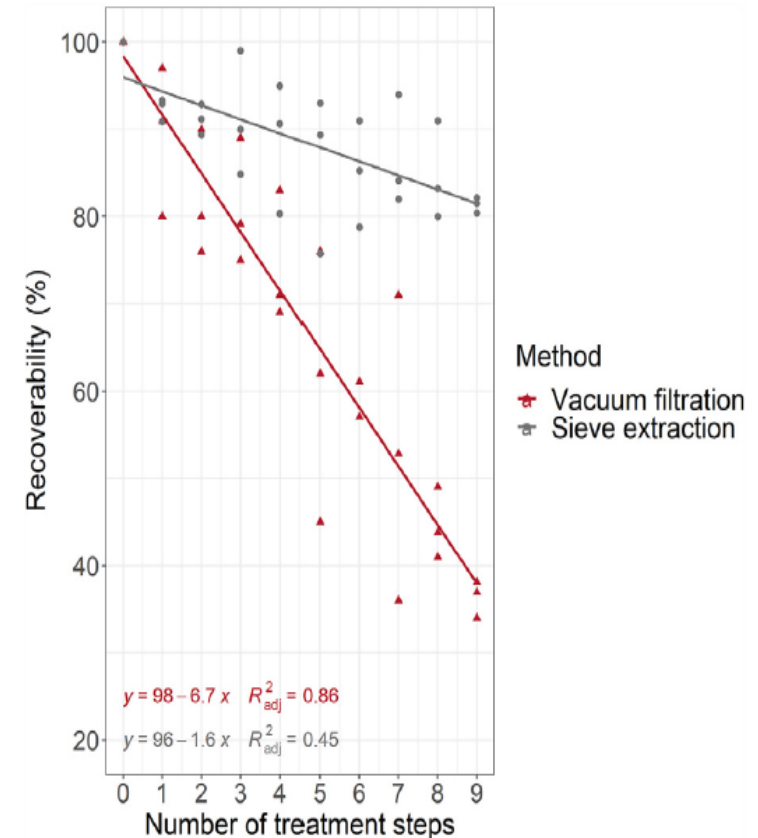


Fig. 4. Recoverability (%) of the 100 polystyrene (PS) beads depending on extraction method (vacuum filtration and sieve extraction) during nine consecutive treatment steps.

Chemical treatment

Physical separation

For water samples oxidation with H_2O_2 (30%) for 2 days at room temperature

For sediment samples no oxidation was made, density separation with NaI (1.8 g/cm^3) repeated for three times

Sieving through 300 μm and 10 μm metal filters

Substrate choice

Sample properties

(Un)known compound

Reflectiveness

Thickness

Roughness

Compatibility

Substrate choice

Depth of analysis

Entire sample

Subsample

Substrate choice

Deposition method

Filter

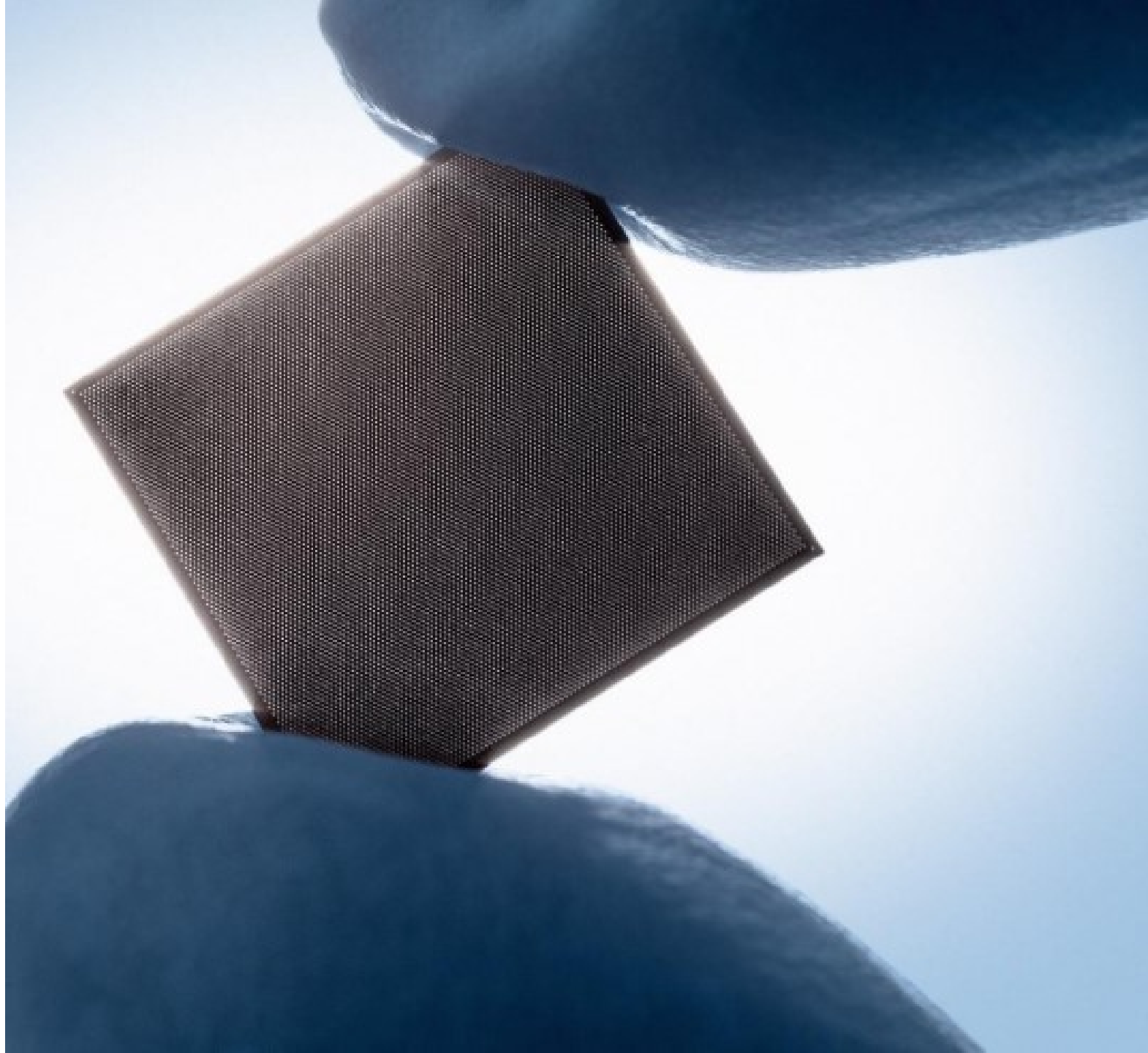
Smear

Compacting disc

Liquid evaporation

Substrate for FTIR imaging

- Silicon filter (transmission) for size $< 300\text{ }\mu\text{m}$ - u FTIR Imaging (Size $11 \times 11\text{ mm}^2$, Si-thickness: $\sim 170 - 230\text{ }\mu\text{m}$, pore size $10\text{ }\mu\text{m}$ in diameter)
- Optical ZnSe window (transmission) for size $> 300\text{ }\mu\text{m}$ - measuring a spectra from particle (point mode)



Why silicon filter?

- Silicon filter can be used to perform both transmission and reflectance analysis in range 500- 4000
- The only negative points about the silicon are the relative cost and the “non-standard” sizes (rectangular dimensions) that are not directly compatible with standard filtration systems



u-FTIR Spotlight 400 (Perkin Elmer)

- ✓ Particle information
- ✓ Material type per particle
- ± Estimated mass
- Need for substrate in lower size range
- Rubber identification difficult



Scans interpretation

For particles $> 300 \mu\text{m}$ -
comparing spectra with in-house
built library of polymers



For particles $< 300 \mu\text{m}$ - scanning
the whole filter and interpreting
using siMPle programme

Microplastic Identification Using siMPle Software: Summary of Methodology and Optimization

- **Software Used:** Polymer identification was performed using the **siMPle** program (classic 2020 version), with the **AAU pipeline** and derivative-based analysis (Pearson weights: 0/1/1).
- **Method Strengths:**
 - Transparent and **robust approach**, requiring no reference spectra for natural materials
 - **Good reproducibility**
- **Limitations and Considerations:**
 - Polymer **Threshold Values (TVs)** need to be carefully optimized to balance **false positives (~5%)** and false negatives
 - Even with optimization, **some MPs may be underreported** due to spectral/matrix limitations.

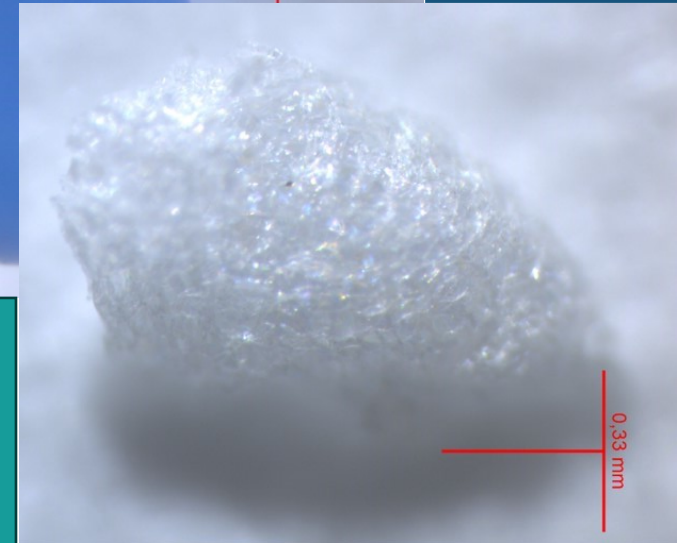
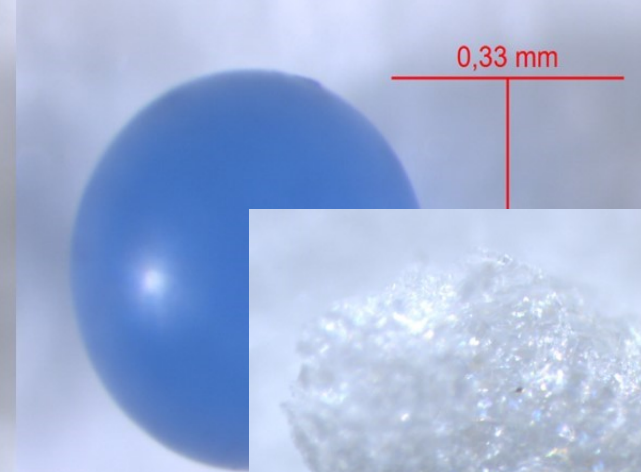
Microplastic Identification Using siMPle Software: Summary of Methodology and Optimization

- **Quality Control:**

- Essential step – **manual visual QC** of MPs is more reliable than automated checks
- Doubtful identifications were cross-checked using an additional spectral database (Open Specy)

- **Conclusion:**

- This method is **not perfect**, but with manual QC and optimized settings, it yields **plausible and conservative** estimates of MP presence
- **Standardization** of spectral analysis methods is essential for cross-study comparability



natalja.buhhalko@taltech.ee

